

Mouse Complement Factor D ELISA Kit

Vertrieb:

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Hinweis/Note:

Der Packungsbeileger dient nur als erste Information. Der relevante Packungsbeileger liegt der Ware bei.

The datasheet is only a first information.

The relevant datasheet is included with the product.

For any questions regarding troubleshooting or performing the assay, please contact our support team at support@assaypro.com.

Thank you for choosing Assaypro.

Assay Summary

Add 50 µl of standard/samples per well. Incubate 2 hours.



Wash, then add 50 µl of biotinylated antibody per well. Incubate 1 hour.



Wash, then add 50 µl of SP per well. Incubate 30 minutes.



Wash, then add 50 µl of Chromogen Substrate per well. Incubate 10 minutes.



Add 50 μ l of Stop Solution per well. Read at 450 nm immediately.

Assay Template

	1	2	3	4	5	6	7	8	9	10	11	12
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AssayMax Mouse Complement Factor D ELISA Kit

Catalog No. EMF7701-1
Sample Insert/Reference Only

Introduction

Complement Factor D (FD), also known as adipsin, is a serine protease secreted by adipocytes into the bloodstream and a component of the alternative complement pathway. It is a 24 kDa single chain glycoprotein with 222 amino acids and has a high level of expression in fat (1-3). FD cleaves factor B bound to C3b, generating the alternative pathway C3 convertase C3bBb and releasing the Ba fragment. Human FD plays a major role in humoral suppression of infectious agents. The deficiency of FD increases susceptibility to infections and their recurrence (4).

Principle of the Assay

The AssayMax Mouse Complement Factor D (FD) ELISA kit is designed for detection of mouse FD in plasma, serum, urine, and cell culture samples. This assay employs a quantitative sandwich enzyme immunoassay technique that measures mouse FD in less than 4 hours. A polyclonal antibody specific for mouse FD has been pre-coated onto a 96-well microplate with removable strips. Mouse FD in standards and samples is sandwiched by the immobilized antibody and the biotinylated polyclonal antibody specific for mouse FD, which is recognized by a streptavidin-peroxidase conjugate. All unbound material is then washed away and a peroxidase enzyme substrate is added. The color development is stopped and the intensity of the color is measured.

Caution and Warning

- Prepare all reagents (working diluent buffer, wash buffer, standards, biotinylated antibody, and SP conjugate) as instructed, prior to running the assay.
- Prepare all samples prior to running the assay. The dilution factors for the samples are suggested in this protocol. However, the user should determine the optimal dilution factor.
- Spin down the SP conjugate vial and the biotinylated antibody vial before opening and using contents.
- This kit is for research use only.
- The kit should not be used beyond the expiration date.

• The Stop Solution is an acidic solution.

Reagents

- Mouse Complement Factor D Microplate: A 96-well polystyrene microplate (12 strips of 8 wells) coated with a polyclonal antibody against mouse FD.
- **Sealing Tapes:** Each kit contains 3 precut, pressure sensitive sealing tapes that can be cut to fit the format of the individual assay.
- **Mouse Complement Factor D Standard:** Mouse FD in a buffered protein base (4 ng, lyophilized).
- **Biotinylated Mouse Complement Factor D Antibody (50x):** A 50-fold concentrated biotinylated polyclonal antibody against mouse FD (140 μl).
- **EIA Diluent Concentrate (10x)**: A 10-fold concentrated buffered protein base (30 ml).
- Wash Buffer Concentrate (20x): A 20-fold concentrated buffered surfactant (30 ml, 2 bottles).
- Streptavidin-Peroxidase Conjugate (SP Conjugate): A 100-fold concentrate (80 μl).
- **Chromogen Substrate**: A ready-to-use stabilized peroxidase chromogen substrate tetramethylbenzidine (8 ml).
- **Stop Solution**: A 0.5 N hydrochloric acid to stop the chromogen substrate reaction (12 ml).

Storage Condition

- Store components of the kit at 2-8°C or -20°C upon arrival up to the expiration date.
- Store SP Conjugate and biotinylated antibody at -20°C.
- Store Microplate, Diluent Concentrate (10x), Wash Buffer, Stop Solution, and Chromogen Substrate at 2-8°C.
- Unused microplate wells may be returned to the foil pouch with the desiccant packs and resealed. May be stored for up to 1 month in a vacuum desiccator.
- Diluent (1x) may be stored for up to 1 month at 2-8°C.
- Store standard at 2-8°C before reconstituting with diluent and at -20°C after reconstituting with diluent.

Other Supplies Required

- Microplate reader capable of measuring absorbance at 450 nm.
- Pipettes (1-20 μl, 20-200 μl, 200-1000 μl and multiple channel).
- Deionized or distilled reagent grade water.

Sample Collection, Preparation and Storage

- **Plasma:** Collect plasma using one-tenth volume of 0.1 M sodium citrate as an anticoagulant. Centrifuge samples at 3000 x g for 10 minutes and use supernatants. Dilute samples 1:10000 with EIA Diluent and assay. The undiluted samples can be stored at -20°C or below for up to 3 months. Avoid repeated freeze-thaw cycles. (EDTA or Heparin can also be used as an anticoagulant.)
- **Serum**: Samples should be collected into a serum separator tube. After clot formation, centrifuge samples at 3000 x g for 10 minutes and remove serum. Dilute samples 1:10000 into EIA Diluent and assay. The undiluted samples can be stored at -20°C or below for up to 3 months. Avoid repeated freeze-thaw cycles.
- **Urine:** Collect urine using sample pot. Centrifuge samples at $800 \times g$ for 10 minutes and assay. Dilute samples 1:10 into EIA Diluent or within the range of 5x 50x. The undiluted samples can be stored at -20°C or below for up to 3 months. Avoid repeated freeze-thaw cycles.
- **Cell Culture Supernatants:** Collect cell culture media and centrifuge at 3000 x g for 10 minutes at 4°C to remove debris. The samples can be stored at -20°C or below. Avoid repeated freeze-thaw cycles.

Reagent Preparation

- Freshly dilute all reagents and bring all reagents to room temperature before use.
- **EIA Diluent Concentrate (10x):** If crystals have formed in the concentrate, mix gently until the crystals have completely dissolved. Dilute the EIA Diluent Concentrate 1:10 with reagent grade water. Store for up to 1 month at 2-8°C.
- Standard Curve: Reconstitute the 4 ng of Mouse Complement Factor D Standard with 2 ml of EIA Diluent to generate a stock solution of 2 ng/ml. Allow the standard to sit for 10 minutes with gentle agitation prior to making dilutions. Prepare duplicate or triplicate standard points by serially diluting the stock solution (2 ng/ml) 1:2 with equal volume of EIA Diluent to produce 1, 0.5, 0.25, 0.125, 0.063, and 0.031 ng/ml solutions. EIA Diluent serves as the zero standard (0 ng/ml). Any remaining solution should be frozen at -20°C and used within 30 days.

Standard Point	Dilution	[Mouse FD] (ng/ml)
P1	Standard (2 ng/ml) + 1 part EIA Diluent	1.000
P2	1 part P1 + 1 part EIA Diluent	0.500
P3	1 part P2 + 1 part EIA Diluent	0.250
P4	1 part P3 + 1 part EIA Diluent	0.125
P5	1 part P4 + 1 part EIA Diluent	0.063
P6	1 part P5 + 1 part EIA Diluent	0.031
P7	EIA Diluent	0.000

- **Biotinylated Mouse Complement Factor D Antibody (50x):** Spin down the antibody briefly and dilute the desired amount of the antibody 1:50 with EIA Diluent. Any remaining solution should be frozen at -20°C.
- Wash Buffer Concentrate (20x): If crystals have formed in the concentrate, mix gently until the crystals have completely dissolved. Dilute the Wash Buffer Concentrate 1:20 with reagent grade water.
- **SP Conjugate (100x):** Spin down the SP Conjugate briefly and dilute the desired amount of the conjugate 1:100 with EIA Diluent. Any remaining solution should be frozen at -20°C.

Assay Procedure

- Prepare all reagents, working standards and samples as instructed.
- Remove excess microplate strips from the plate frame and return them immediately to the foil pouch with desiccants inside. Reseal the pouch securely to minimize exposure to water vapor and store in a vacuum desiccator.
- Add 50 μ l of Mouse Complement Factor D Standard or sample per well. Cover wells with a sealing tape and incubate for 2 hours. Start the timer after the last sample addition.
- Wash five times with 200 μ l of Wash Buffer manually. Invert the plate each time and decant the contents; hit 4-5 times on absorbent material to completely remove the liquid. If using a machine, wash six times with 300 μ l of Wash Buffer and then invert the plate, decanting the contents; hit 4-5 times on absorbent material to completely remove the liquid.
- Add 50 μ l of Biotinylated Mouse Complement Factor D Antibody to each well and incubate for 1 hour.
- Wash the microplate as described above.
- Add 50 μ l of Streptavidin-Peroxidase Conjugate per well and incubate for 30 minutes. Turn on the microplate reader and set up the program in advance.
- Wash the microplate as described above.
- Add 50 μl of Chromogen Substrate per well and incubate for approximately 10 minutes or till the optimal blue color density develop.

- Gently tap the plate to ensure thorough mixing and break the bubbles in the well with pipette tip.
- Add 50 μ l of Stop Solution to each well. The color will change from blue to yellow.
- Read the absorbance on a microplate reader at a wavelength of 450 nm immediately. If wavelength correction is available, subtract readings at 570 nm from those at 450 nm to correct optical imperfections. Otherwise, read the plate at 450 nm only. Please note that some unstable black particles may be generated at high concentration points after stopping the reaction for about 10 minutes, which will reduce the readings.

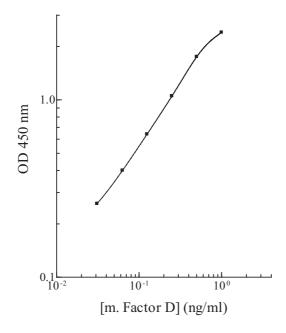
Data Analysis

- Calculate the mean value of the duplicate or triplicate readings for each standard and sample.
- To generate a standard curve, plot the graph using the standard concentrations on the x-axis and the corresponding mean 450 nm absorbance on the y-axis. The best-fit line can be determined by regression analysis using log-log or four-parameter logistic curve-fit.
- Determine the unknown sample concentration from the Standard Curve and multiply the value by the dilution factor.

Standard Curve

 The curve is used for illustration only. A standard curve should be generated each time the assay is performed.

M. Factor D Standard Curve



Performance Characteristics

- The minimum detectable dose of mouse FD is typically ~ 0.03 ng/ml.
- Intra-assay and inter-assay coefficients of variation were 5.0 % and 7.2 % respectively.

Linearity

	Average Percentage of Expected Value			
Sample Dilution	Plasma	Serum		
1:5000	93%	89%		
1:10000	99%	97%		
1:20000	101%	107%		

Recovery

Standard Added Value	0.05 – 0.5 ng/ml
Recovery %	84-113%
Average Recovery %	98%

Cross-Reactivity

Species	% Cross Reactivity
Canine	None
Bovine	None
Monkey	None
Mouse	100%
Rat	None
Swine	None
Rabbit	None
Human	None

References

- (1) Niemann MA et al. (1984) Biochemistry 23(11):2482-2486
- (2) Rosen BS et al. (1989) Science 244(4911):1483-1487
- (3) White RT et al. (1992) J Biol Chem. 267(13):9210-9213
- (4) Sprong T et al. (2006) Blood 107(12):4865-4870

Version 1.0

Related Products

 EF7001-1 AssayMax Human Complement Factor D ELISA Kit (Plasma, Serum, Saliva, Milk, and Cell Culture samples)

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